IBC Meeting Minutes

Chair- Ken Bondioli
Research Safety- Abigail Fish
Thursday, July 16, 2026
1:30 pm via Zoom

Institutions *Louisiana State University Agricultural and Mechanical College (A&M)*
Louisiana State University Ag Center

IBC Members			
	Ken Bondioli	LSU Ag Center	Chair, Animal Expert
	Abigail Fish	LSU A&M	BSO, Administrator, Voting Contact
	Sarah Keeton	LSU A&M	BSO
	Sue Hagius	LSU Ag Center	Animal Expert, Lab Rep
	Michael Hooks	LSU A&M	Member
	Jong Ham	LSU Ag Center	Plant Expert
	Christy White	Pennington Biomedical Research Center	Non-Voting Member
	Jeff Davis	LSU Ag Center	Plant and Insect Expert
	Niranjan Baisakh	LSU Ag Center	Plant Expert
	William Doerrler	LSU A&M	Member
	Ramanuj Lahiri	National Hansen's Disease Program	Member
	Michelle Dennis	Our Lady of the Lake Hospital	Local Non-Affiliated Member
	Brent Stanfield	LSU A&M	Member
	Ryoichi Teruyama	LSU A&M	Member
	James Bush	Blue Cross Blue Shield of Louisiana	Local Non-Affiliated Member

Members Present: Ken Bondioli, Abigail Fish, Sarah Keeton, Sue Hagius, Michael Hooks- proxy Samantha Clark, Ryoichi Teruyama, William Doerrler, and Niranjan Baisakh.

Members Absent: Christy White, Michelle Dennis, Jeff Davis, Brent Stanfield, Jong Ham, and James Bush.

Others Present:	Jonathan Richards	Associate Professor, Plant Pathology and Crop Physiology
	Shavahn Loux	Assistant Professor, School of Animal Sciences

Call to Order: 1:32 pm

Approval of Minutes from: Meeting minutes for June are not yet available. Minutes will be completed and voted on at the August IBC meeting
Motion Made by:
Seconded by:
Abstaining:

Business and Call for New Business

Rebecca Christofferson has been removed as a member of the IBC because she is no longer with the University.

New IBC Registrations and Amendments for Review

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26026 (On Hold)	Jonathan Richards	Plant Pathology and Crop Physiology	07/06/2026	Functional Genomics and Molecular Biology of Fungal Pathogen-Plant Interactions	Niranjan Baisakh	William Doerrler

Project Overview:	The overall goal of this project is to identify and characterize the molecular mechanisms that contribute to virulence in *Cercospora* species that cause disease in soybean and rice. The research will focus on identifying predicted fungal effector proteins and determining how these proteins may promote pathogen fitness by altering host physiological processes, inducing plant cell death or necrosis, or otherwise contributing to disease development.
Risk Assessment and Discussion:	This project poses a low biosafety risk associated with handling plant-pathogenic *Cercospora* species, recombinant DNA constructs, transformed fungal isolates, and infected soybean or rice plants. The primary concern is the accidental dispersal or environmental release of fungal spores, inoculum, genetically modified isolates, or infected plant material. Potential hazards include exposure to spores or aerosols during fungal culture, blending, filtration, centrifugation, inoculum preparation, spraying, and handling of infected plants and soil. Additional risks include contact with recombinant bacterial, yeast, and fungal cultures; purified fungal proteins that may induce plant tissue necrosis; and naturally produced cercosporin from *C. janseana* and *C. cf. flagellaris*. These risks are managed using BSL-1 and BL2-P containment, appropriate PPE, Class II biosafety cabinets, enclosed mist chambers, restricted access, sealed transport containers, secure culture storage, routine inventory controls, bleach disinfection, and autoclaving of cultures, infected plants, soil, and contaminated waste. The proposed genetic modifications are not expected to increase fungal virulence and will not involve interspecies gene transfer, novel effector mutations, or modification of cercosporin biosynthesis. With these precautions in place, the project is considered low risk and appropriately contained under BSL-1 and BL2-P practices
NIH Guidelines:	Section III-E-2-b-(3)
Biosafety Level:	BSL-1 and BL2-P

Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-1 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-1 and BL2-P pending receipt of modifications**
Motion made by:	Niranjan Baisakh
Seconded by:	William Doerrler
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:

- Section B. Project Description.
  - Procedures and Methods. Please clarify where fungal transformations are done and if that work takes place inside a BSC. Please add a statement on secure transport between the lab where the BSC is located and your lab. Objective 2. Please indicate that genes of interest are the genes identified under objective 1. Please indicate that the mist chamber is located in the growth room. Please clarify whether an autoclave is located in the greenhouse; if not, please add a statement about transporting waste back to the lab. Please indicate that the growth chamber is located in the greenhouse.
- Section C. Risk Evaluation.
  - Biosafety. Please indicate what PPE is worn when working in the greenhouse.
  - Biosecurity. Please state that you will use a final concentration of 10% bleach for a minimum contact time of 30 minutes prior to liquid waste disposal.
- Section I. Plant Pathogens.
  - Please update the total volume of stock culture. Source of pathogen. Please indicate which lab, department or PI provides the pathogens.
- Section N. Safety.
  - Administrative Controls. Disinfection/Decontamination. Under other, please describe how plant material is decontaminated before disposal.
  - Engineering Controls. Biosafety Cabinet. Please indicate if the BSC is located in a containment suite or in the general lab area.

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26027 (On Hold)	Shavahn Loux	Animal Sciences	07/07/2026	Microbial Pathogens Associated with Equine Disease	Sarah Keeton	Ryoichi Teruyama

Project Overview:	The overall goal of this project is to better understand the bacteria associated with equine reproductive disease, particularly focal mucoid placentitis and suspected bacterial endometritis. The research will evaluate potential environmental sources of these organisms, characterize their genetic and proteomic features, examine interactions between the bacteria and equine reproductive tissues, and assess the antimicrobial susceptibility of isolates recovered from affected mares.
Risk Assessment and Discussion:	This project poses a moderate biosafety risk associated with collecting and processing environmental samples, equine reproductive specimens, bacterial cultures, and tissues potentially containing unidentified microorganisms. Organisms associated with equine placentitis or endometritis may include opportunistic or zoonotic bacteria, and the microbial composition of pasture, soil, plant, water, uterine, and placental samples may not be fully known at the time of collection. Potential hazards include exposure during sample collection, transport, culture, identification, antimicrobial susceptibility testing, and host–bacterial interaction studies. Personnel may be exposed through direct contact with contaminated materials, accidental inoculation, splashes, droplets, or aerosols generated during culture manipulation, centrifugation, vortexing, homogenization, or preparation of bacterial suspensions. Additional risks include handling antimicrobial agents and other laboratory reagents used during bacterial characterization and susceptibility testing. These risks are managed under BSL-2 practices using appropriate PPE, controlled sample transport, aerosol-reduction procedures, biosafety cabinets for procedures that may generate aerosols, surface and equipment decontamination, secure culture storage, and proper treatment and disposal of biological waste. With these precautions in place, the project is considered moderate risk and appropriately contained under BSL-2 practices.
NIH Guidelines:	Not Applicable
Biosafety Level:	BSL-2

Training Requirements: All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs

IBC Vote: **Approved at BSL-2 pending receipt of modifications**

Motion made by: Sarah Keeton

Seconded by: Ryoichi Teruyama

Abstaining: None

Conflicts of Interest: None

Requested Modifications:

[redacted]
Confidential - Not subject to NIH Guidelines

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26030 (Renewal)	Adam Bohnert	Biological Sciences	06/24/2026	Reversal of Age-Related Mitochondrial Damage in Caenorhabditis elegans Germline	Ryoichi Teruyama	Niranjan Baisakh

Project Overview:	The overall goal of this project is to determine how mitochondrial turnover and quality control are regulated in the *C. elegans* germline and how these processes change with parental age. The study will use mitophagy sensors and live imaging to evaluate mitochondrial renewal during germline development and oocyte maturation, including the effects of sperm signaling, lysosome activity, CDC-48/VCP/p97 function, aging, oxidative damage, and metabolic state.
Risk Assessment and Discussion:	This project poses a low biosafety risk associated with handling *C. elegans*, recombinant DNA constructs, plasmids, and genetically modified worm strains expressing mitochondrial and mitophagy sensors. *C. elegans* is nonpathogenic to humans, and the proposed genetic modifications are intended to visualize normal mitochondrial turnover and quality-control processes rather than alter pathogenicity or host range. Potential hazards include accidental exposure to recombinant materials, bacterial cultures used for cloning or worm maintenance, and minor risks associated with routine laboratory procedures such as microinjection, centrifugation, pipetting, and fluorescence imaging. These risks are managed under BSL-1 practices using appropriate PPE, standard microbiological techniques, careful sharps handling, secure storage of plasmids and strains, surface decontamination, and autoclaving of biological waste. No significant environmental release or biosecurity concerns are anticipated. With these precautions in place, the project is considered low risk and appropriately contained under BSL-1 practices.
NIH Guidelines:	Section III-D-4-a
Biosafety Level:	BSL-1 and BL1-N
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-1 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-1 and BL1-N pending receipt of modifications** Motion made by: Ryoichi Teruyama Seconded by: Niranjan Baisakh Abstaining: None Conflicts of Interest: None

Requested Modifications:

- Section A. Project Information.

    - Locations. Please add the building for the shared instrument facility
    - Room numbers. Please add the room number for the autoclaves and fluorescent imaging room.
    - Personnel. Please list the BSL-1 EHS-required safety training courses for all staff and the PI.
- Section B. Project Description.
    - Project Goals. Please state an overall project goal and indicate why CDC48/VCP/p97 are important to this work.
    - Procedures and Methods. Please indicate what work takes place in what lab. Please describe how you generate the fluorescent constructs. Please indicate whether you plan to express any genes other than the fluorescent reporters. Please spell out all acronyms, such as ROS and UTR, the first time you use them. Please briefly describe standard worm microinjection techniques. Please elaborate on transgene insertion into landing pads and describe how transgenic lines are identified. Please indicate if you plan to cross deletion strains with fluorescent marker strains. Please indicate what type of E. coli is used and briefly describe bacterial culture techniques. Please indicate if you have specific gene targets for germline development and autophagy disruption. Please describe FZO and mRUBY strains. Please briefly describe genes VCP, p97, and CDC48. Please add the room number for imaging. Please briefly describe the live germline staining approach and include a statement on secure transport if the material is moved between labs. Please add a lab coat and safety glasses for the required PPE.
- Section C. Risk Evaluation
    - Biosafety. Please add the use of safety glasses for required PPE and describe relevant training, including EHS required and lab-specific training. Please add a statement on aerosol containment.
    - Biosecurity. Please indicate how you discard liquid biological waste and worms. Please spell out AMAC
- Section F. Recombinant DNA.
    - Please ensure this entire section includes the ski-lodge integration system and the RNAi information including genes expressed, plasmids, other vectors, experimental hosts, etc.
- Section G. Transgenic Animals
    - Please ensure items 5 through 10 capture information for the ski-lodge integration plasmids and FZO and mRUBY constructs, and those constructs are listed in the attached Excel file. Item 8 indicates that some transgenic animals will experience a loss of function. Please describe others.
- Section N. Safety.
    - Administrative Controls. Stock Cultures. Please list the room number for the -80 freezer.

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26031 (Renewal)	Mariano Carossino	Pathobiological Sciences	07/02/2026	PARTNERSHIP: Virus Replication Kinetics and Risk of Spillback of North American Cervid-Adapted SARS-CoV-2 Variants in Deer and Wildlife	William Doerrler	Sue Hagius

Project Overview:	The overall goal of this project is to evaluate the replication, host range, pathogenicity, transmission potential, and risk of wildlife reservoir establishment of North American white-tailed deer–adapted SARS-CoV-2 variants. The study will use a white-tailed deer-adapted virus containing naturally occurring deer-derived spike sequences, compare their replication and neutralization characteristics in ruminant, rodent, and human cell systems, and assess host susceptibility and transmission in deer mice, white-footed mice, and white-tailed deer. These studies will help determine the role of adaptation in deer and its impact on transmission within wildlife populations and potential spill back into other animals or humans.
Risk Assessment and Discussion:	This project poses a high biosafety risk associated with the generation and handling of infectious recombinant SARS-CoV-2 containing naturally occurring spike sequences identified in white-tailed deer. The primary hazards at LSU are occupational exposure to infectious virus, exposure to infected rodents, tissues, and clinical samples, and accidental release from containment. Potential exposure may occur through aerosols, droplets, splashes, mucous membrane contact, accidental inoculation, or contact with contaminated surfaces during virus propagation, cell culture studies, rodent procedures, specimen collection, necropsy, and handling of infectious samples. The recombinant viruses may exhibit differences in replication, pathogenicity, or transmissibility; however, the spike sequences being evaluated occur naturally and are not being engineered to create novel mutations. These risks are managed at LSU through BSL-3 and ABSL-3 containment for infectious-virus and rodent studies, appropriate PPE and respiratory protection, restricted access, validated inactivation procedures, and secure storage and inventory controls. White-tailed deer infections will be conducted separately at the Kansas State University BSL-3Ag facility, with only collected samples shipped to LSU for analysis. With these controls in place, the LSU portion of the project is considered high risk and appropriately contained under BSL-3 and ABSL-3 practices.
NIH Guidelines:	Section III-D-1-b, Section III-D-2-a, Section III-D-4-b and Section III-E-1
Biosafety Level:	BSL-2, BSL-3, ABSL-2, ABSL-3, BL2-N, and BL3-N
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-3 and ABSL-3 training, as applicable to their assigned duties, in accordance with LSU's Environmental Health and Safety (EHS), Institutional Biosafety Committee (IBC), and high-containment program requirements. Personnel handling recombinant SARS-CoV-2, infected animals, infectious samples, or inactivated materials must also complete all required agent-specific, respiratory protection, occupational health, and laboratory-specific training before beginning work. All training must be refreshed as required by LSU policies, approved SOPs, and high-containment facility requirements.
IBC Vote:	**Approved at BSL-2, BSL-3, ABSL-2, ABSL-3, BL2-N and BL3-N pending receipt of modifications** Motion made by: William Doerrler

Seconded by:	Sue Hagius
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:

- Section A. Project Information.
  - Locations. Please remove the room number for the ultra-centrifuge if it is not being used.
  - Personnel. Please list BSL-3 EHS-specific training under specific training for all personnel, including the PI.
- Section B. Project Description.
  - Project Goals. Please shorten this section and rewrite in layman's terms. Please include a statement on the importance of ACE2
  - Procedures and Methods. Please move the first paragraph to the project goals. Please indicate how you obtain primary human nasal epithelial cells and indicate what cells are used for what procedures, including CRFK cells. Please indicate what takes place in what lab. Please state that flow cytometry is done on fixed cells.
- Section C. Risk Evaluation.
  - Biosecurity. Please add a statement on secure transport between labs.
- Section D. Project Units. Please indicate that IACUC protocol will be renewed as needed.
- Section F. Recombinant DNA.
  - NIH Guidelines. Please add Section III.D-4-b.
  - Please add MDOK cells if relevant.
- Section J and K. Human and Animal Pathogens.
  - Method of concentration. Please uncheck other.
- Section M. Human and Primate Blood, Body Fluids, or Tissues
  - Please check primary culture
- Section N. Safety.
  - Disinfection/Decontamination. Please indicate what is used for routine cleaning and spills.

Upcoming Meetings:	August 13, 2026 @1:30 pm via Zoom
Adjourned:	2:28 pm